

Title	Legionella Specimen Collection and Transport
Specimen Requirements	<p>Acceptable for PCR:</p> <p>Nasopharyngeal swabs</p> <p>Acceptable for Culture:</p> <p>Sputum</p> <p>Bronchial washings</p> <p>Bronchial or tracheal aspirates (30-50 mL recommended)</p> <p>Bronchial alveolar lavage (BAL) fluids (30-50 mL recommended)</p> <p>Sterile body fluids (CSF, pericardial, peritoneal, pleural, etc.; 5 mL recommended)</p>
Sampling Materials	<ul style="list-style-type: none"> • Transport media: Appropriate bacterial or viral transport media available commercially. Check package insert to confirm if media requires refrigeration. • Swab: Swab material must be synthetic (<i>i.e.</i> rayon, polyester, or Dacron). Calcium alginate or charcoal-impregnated swabs may not be used. Wood-shaft and dry swabs are not accepted. • Sterile containers and collection materials are required for aspirate, wash, BAL, and sputum specimens. • Use cold packs or dry ice if media requires refrigeration or specimen is transported frozen. • Ship boxes/containers with appropriate shipping labels. • •
Procedural Notes	<ul style="list-style-type: none"> • • Label the specimen tube with the patient's first/last name, date of birth, and date of collection. • • Check the expiration date on the transport media tube to ensure product is acceptable and will continue to be acceptable once received at the laboratory. • • After collection, all specimens should be stored refrigerated (2-8°C) until shipped; specimens should be shipped for receipt within 5 days. If longer storage is required, keep specimens at -70°C. Avoid freeze-thaw cycles. • <p>Special Instructions for Specimen Collection</p> <p>NP Swab: Insert Dacron-tipped swab through the nostril into the nasopharynx until tip reaches distance equivalent to that from the ear to the nostril of the patient. Rotate swab several times, remove carefully, and place swab into the VTM tube.</p> <p>Nasal Wash: Place several milliliters of sterile saline into nostrils while patient's head is tilted back. Bring patient's head forward and catch saline flowing from the nostrils into a small container. Pour 1-2mL of specimen into the VTM tube.</p> <p>Sputum: Satisfactory quality implies the presence of mucoid or mucopurulent material. Sputum specimen should have a volume of 3-5mL, although smaller quantities are acceptable if the quality is satisfactory.</p>

Shipping Instructions	Please contact the Clinical Microbiology Director with questions: 317-921-5894 or sblosser@isdh.in.gov. Submit a completed "Reference Submission Form" with each specimen to be submitted.
Reporting and TAT	